Supramolecular Device for Artificial Photosynthetic Mimics As Helix-Mediated Antenna/Reaction Center Ensemble

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ABSTRACT

We have developed a novel integrated supramolecular device for a photosynthetic antenna/reaction center (RC) model based on a helical amylose, which plays an important role as the host for cyanine dye J-aggregation onto the helical surface and also for inclusion of a D−**A chain chromophore inside the helical cavity, where the J-aggregates function as an array of photoreceptor antenna that funnel excitation across the helix to the chromophore.**

In nature, photosynthetic bacteria have evolved intricate selfassemblies of chlorophyll and carotenoid aggregates onto proteins as scaffolds. Most chlorophylls serve as lightharvesting antenna (LH-I and LH-II) that capture the sunlight and funnel the electronic excitation cascading into the photosynthetic reaction center (RC) ,¹ where the photon energy is converted into electronic energy.

The light-harvesting complexes are circularly arranged as such that LH-I complexes surround the RC and are surrounded in turn by LH-II antenna complexes. It is assumed that the excitation energy is delocalized over the aggregates and excitation transfer takes place effectively between interas well as intracomplexes, achieving the quantum yield of near unity at $RC¹$.

There have been various types of covalently linked, multichromophoric systems reported² for artificial photosynthetic mimics, but little is known for a supramolecularly integrated device as an antenna/RC ensemble. We were interested in developing a helical supramolecular device mimicking a photosynthetic antenna. We envisioned Jaggregates of a cyanine dye can play an important role as an antenna array due to the very large absorption crosssection and efficient exciton migration over many molecular aggregates.3 Interestingly, when they are bound to a rigid helical surface, and an acceptor is incorporated inside the helix, the device configuration (see the Abstract graphic)

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uniquely resembles the circular arrangement of a photosynthetic antennas, such that the photoexcited energy of the J-aggregates (antennas) in the periphery is directed onto the acceptor (RC) in the center, thereby amplifying the energytransfer quenching (Figure 1).

Figure 1. Chemical structures of the supramolecular device components: carboxymethyl amylose (CMA) and cyanine dyes.

First, we have investigated the possible J-aggregation formation of a cyanine dye (cyanine-1) in the presence of carboxymethyl amylose (CMA) and found that the cyanine J-aggregation occurs onto the CMA at various degrees of carboxymethylation (DS), forming a CMA/cyanine-1 superhelix,⁴ while CMA itself is a random-coil in solution due to the electrostatic repulsion. The super-helix formation was proven by an extraordinarily large induced circular dichroism (CD) and a large enhancement $(> \times 10^2)$ of fluorescence
intensity ($\Phi_0 = 0.43$) of the cyanine dye Lagoregates⁴ intensity ($\Phi_f = 0.43$) of the cyanine dye J-aggregates.⁴

From the distance effect consideration⁵ on the excitation transfer (from the antenna to an energy/electron-acceptor), the inside of the super-helix is the best accessible site for an acceptor, which is therein circularly surrounded by the J-aggregate antenna array.

Not only that, under this circumstance the chromophore takes advantage⁶ of the helical host, benefiting from aggregation-free, single molecular confinement within the helix, which gives a longer excited-state lifetime.⁷ In a recent communication, we reported that when a flexible-chain, electron donor-acceptor $(D-A)$ chromophore is helically encapsulated, the electron transfer (eT) prevails along the helical axis with a clear distance dependence. By contrast,

the eT of the encapsulation-free counterpart exhibits no distant dependence and a very poor efficiency due to a selfquenching resulting mainly from aggregation and/or solvent deactivation of chromophores.7 In addition, it is worth noting that the helical encapsulation of a chromophore is an effective means to develop one-dimensional self-assembly,⁸ since no self-assembly occurs with the encapsulation-free counterpart, suggesting that lateral interactions of helices including H-bonding promote the surface binding.

In this paper, we report a unique supramolecular nanodevice for an artificial photosynthetic antenna/RC ensemble based on CMA and two-component dyes (cyanine dye and ^D-A chromophore), where CMA is indispensable not only for imparting the functional role to the antenna but also accommodating a chromophore (DASP-C₂₂ or DASP- $_6$ V₁₂)^{6,7} into the cavity of the super-helix (Figure 2). The device is prepared by integration of two competitive (**A**, **B**) but sequentially controlled self-organization processes (**B**), namely, J-aggregation of a cyanine dye onto CMA surface, forming super-helix, and the subsequent treatment of DASP-chromophore for encapsulation with the super-helix. This procedure is critical for the completion of the device (see the Supporting Information, S-1), particularly with CMA of a high DS (> 0.1). The occurrence of the processes was verified by UV/vis and CD spectra of individual chromophores^{4,6,7} and their fluorescence intensity changes due to excitation transfer (from the antenna) to $DASP-C_{22}$ (energy-acceptor), along with the concomitant increase in the DASP- C_{22} emission due to energy-transfer (ET). The transfer efficiency is assessed by changes in the fluorescence intensity in the presence and in the absence of the encapsulated DASP-C₂₂ or DASP- $_6V_{12}$. Also, the time-resolved fluorescence lifetimes of the J-aggregates and the DASP chromophores are measured (see the Supporting Information, S-1) and correlated.

For CMA, it is almost impossible to assume a helical conformation by itself, and even with an extremely low degree (e.g., $DS = 0.06$) of carboxymethylation, the helical encapsulation of DASP, for example, is significantly decreased compared to that of neutral amylose, such that at a higher DS (> 0.1), CMA loses the inclusion capability completely (see the Supporting Information, S-2). However, surprisingly enough, when the higher DS, random-coil CMA is transformed into a super-helix by cyanine J-aggregation, it turns out perfectly capable of encapsulating $DASP-C_{22}$ and even $DASP_{6}V_{12}$; the latter reveals a particularly strong affinity (Figure 2, route **B**) for the highest DS (1.53) CMA super-helix, thereby rendering a remarkable stability, 9 while for the lowest DS (0.06) CMA super-helix, its charge interaction with $DASP_{-6}V_{12}$ is destructive, rendering the encapsulation detrimental.¹⁰ Accordingly, for a low DS $($ 0.1) CMA, the alternative route **A** is preferred. The helical

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⁽⁸⁾ Remarkably, this supramolecule forms self-assembly thin films on various substrates. This will be published elsewhere in detail.

⁽⁹⁾ The supramolecular device prepared using the high DS (1.53) CMA super-helix are very stablesuch that their UV/vis and CD spectra in aqueous solution exhibit no significant changes at least for 1 month.

⁽¹⁰⁾ DASP- $_6V_{12}$ is very cooperative with the high DS (1.53) CMA superhelix for encapsulation. This may be associated with the tight super-helix structure (see the Supporting Information, S-3).

Figure 2. Pictorial representation of supramolecular architecture for the photosynthetic antenna/RC ensemble, where the DASP-viologen (D-A) chain chromophore is encapsulated by the super-helix of CMA/cyanine J-aggregates as antenna. Inset shows a different CD intensity in the cyanine J-aggregates, reflecting the state of the super-helix with CMA ($DS^* = 0.06$).

cyanine/CMA super-structure is already heavily twisted,⁴ but it becomes even more pronounced, as shown in Figure 2 (inset), upon encapsulating a chromophore (DASP- C_{22}).

Figure 3 (left) shows a strong absorption band at 460 nm (a, b, c) due to the cyanine J-aggregation of the super-helix (with 1.53 DS CMA), which is unaffected by the presence (encapsulation) of DASP-C₂₂ or DASP- $_6V_{12}$, and a weak and broad absorption band around 480 nm (d) for the free DASP- $6V_{12}$ in a random-coil CMA (1.53 DS) solution. The peak position of DASP- C_{22} is commonly red-shifted⁶ when encapsulated with amylose. By the same token, with the super-helix encapsulation the absorption band of $DASP_{-6}V_{12}$ (c) is red-shifted to 500 nm from 475 nm of the encapsulation-free counterpart (d). This is reflected in the fluorescence emission bands (see Figure 4 inset and Supporting Information, S-4). The strong J-band as a proof of the super-helix encapsulation (b, c) is manifested by CD spectra (right), 11 in that the induced CD (of the J-aggregates) remains mostly unchanged by treating (encapsulating) $DASP-C_{22}$ (b) and $DASP_{-6}V_{12}$ (c). By contrast, the super-helix based on the lowest DS (0.06) CMA is disrupted (see the Supporting Information, S-3) by the presence of $DASP-6V_{12}$ to monomeric cyanine,¹² losing the CD signal of the helical J-band.

The fluorescence spectra (Figure 4, left) provide conclusive evidence for the super-helical inclusion of DASP- C_{22} , which brings about a sharp fluorescence quenching (ca. 90%) of

Figure 3. Absorption (left) and CD (right) spectra of cyanine J-aggregates/CMA (1.53 DS) super-helix: (a) super-helix alone (with no chromophore encapsulated), (b) $DASP-C_{22}$ encapsulating super-helix, (c) DASP-6V₁₂ encapsulating super-helix, and (d) CMA $(1.53 \text{ DS})/\text{DASP}_{5}V_{12}$ mixture; [cyanine] = 2.3 × 10⁻⁵ M, [CMA (1.53 DS)] = 0.9 × 10⁻³ M, and [DASP-C₂₂] = [DASP-₆V₁₂] = 1.7×10^{-5} M in H₂O.

the J-aggregates at 475 nm, accompanying the concomitant enhancement ($> \times 20$ ^{7a} of DASP fluorescence (at 605 nm) due to ET (details in inset). A similar red-shift (15 nm) is observed for $DASP_{-6}V_{12}$ in the emission band due to the encapsulation (see Supporting Information, S-4 and Note). When $DASP_{-6}V_{12}$ (instead of $DASP-C_{22}$) is encapsulated by the super-helix, the excited-state of the DASP moiety is almost completely quenched by eT of the viologen subunit $({}_6V_{12})$ to a charge-separated state. The overall quenching efficiency along the pathways, from photoinduced ET from

⁽¹¹⁾ There is a close correlation between the absorption and CD spectra (Figure 2) of $DASP_{-6}V_{12}$ in the presence and absence of the super-helix (based on the cyanine J-aggregation with a high DS (1.53) CMA). (12) It is likely that the multiple-charges of DASP- ${}_6V_{12}$ override a relatively weak cyanine J-aggregation with the low DS (0.06) CMA.

Figure 4. (Left) Changes in the fluorescence intensity (at 475 nm, ex at 457 nm) (a) of cyanine J-aggregates/CMA (1.53 DS) superhelix, (b) upon encapsulation of $DASP-C_{22}$ by the super-helix, accompanied by an enhancement at 605 nm (upward arrow), (c) upon encapsulation of DASP-DASP- $_6V_{12}$ by the super-helix, accompanied by decreased intensity (downward broken arrows), and (d) of the CMA (1.53 DS)/DASP- $_6V_{12}$ mixture; [cyanine] = 2.3 \times 10⁻⁵ M, [CMA (1.53 DS)] = 0.9 \times 10⁻³ M, and [DASP-C₂₂] = [DASP- $_6V_{12}$] =1.7 × 10⁻⁵ M in H₂O. (Right) Time-resolved fluorescence lifetimes of the photoexcited species; the lifetimes were detected at 475 nm for τ_1 , τ_2 , and τ_3 and at 605 nm for τ_4 and τ_5 (τ values are shown as an average).

the antenna and the subsequent eT to $DASP_{-6}V_{12}$ is $>90\%$ (Scheme 1).

A time-resolved fluorescence decay profile of the excitedstate species along the transport pathways (above) is illustrated in Figure 4 (right). This demonstrates that when detected at 475 nm, the fluorescence lifetime (τ_1) of the J-aggregates (of the super-helix) is decreased by a factor of 5 due to ET quenching (τ_2) by DASP-C₂₂, and the subsequent eT quenching by the ${}_6V_{12}$ subunit further decreases the lifetime (τ_3) by at least another factor of 3. Such a dramatic shortening of J-aggregates lifetime is a clear indication of ET/eT along the pathways. Separately, when the decay was detected (for eT) at 605 nm (for τ_4 or τ_5), the τ_4 is longer compared to the τ_3 . This is unclear but can be explained by that the DASP excited-state is interfered by a concomitant deactivation process¹³ involving a direct eT to the ${}_6V_{12}$ subunit (in encapsulation) from the excited J-aggregates. Nonetheless, the eT rate is still very fast: $k_e = 1/\tau_{D-A}$ - $1/\tau_{\rm D} = 1.3 \times 10^{10} \text{ s}^{-1}$. However, compared to τ_4 , the
unusually shortened τ_5 (due to the free DASP_r V_{L3} dissolved unusually shortened τ_5 (due to the free DASP- $_6V_{12}$ dissolved in the random-coil CMA solution) is not due to the efficient eT but is attributed to self-quenching^{7b} resulting from aggregations, particularly due to CMA-mediated electrostatic binding of the free $DASP_{-6}V_{12}$ and/or nonradiative deactivation of the polar solvent (Supporting Information, $S-4$ note).¹⁴

In conclusion, we have constructed a unique supramolecular device for an artificial photosynthetic mimic by integration of a functional amylose (a high DS CMA) with two interactive dye molecules; one on the helical surface and the other inside the helix. Cyanine dye J-aggregates complexed with CMA (super-helix) function as an array of photoreceptor antennas, and a donor-acceptor $(D-A)$ chromophore encapsulated by the super-helix functions as reaction center (RC). Steady-state energy/electron transfer (ET/ eT) was investigated throughout the excitation transfer pathways and correlated to their time-resolved lifetimes. From the high transfer efficiency and the stability, we believe that the supramolecular device as an antenna/RC ensemble can be a viable model for artificial photosynthesis.

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Supporting Information Available: Additional spectral data are available. This material is available free of charge via the internet at http://pubs.acs.org.

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⁽¹³⁾ The $6V_{12}$ subunit (inside the super-helix) in proximity to the J-aggregates (on the super-helix surface) can also quench the excitation of the J-aggregates directly, not through DASP subunit, such that τ_4 (involvingDASP to $_6V_{12}$ eT process) becomes longer compared to τ_3 (involving the J-aggregate quenching by DASP subunit).

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